



CERTIFICATION

AOAC Research Institute *Performance Tested Methods*SM

Certificate No.

072601

The AOAC Research Institute hereby certifies the method known as

Elevia 1.1 *Salmonella* spp.

manufactured by

TAAG Technologies

Rio Refugio 9641

Santiago, Chile

This method has been evaluated and certified according to the policies and procedures of the AOAC *Performance Tested Methods*SM Program. This certificate indicates an AOAC Research Institute Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC Research Institute *Performance Tested Methods*SM certification mark on the above-mentioned method for the period below. Renewal may be granted by the Expiration Date under the rules stated in the licensing agreement.

A handwritten signature in black ink, appearing to read "Bradley A. Stawick".

Bradley A. Stawick, AOAC Research Institute Senior Director

Issue Date

July 10, 2026

Expiration Date

December 31, 2026

METHOD NAMEElevia 1.1 *Salmonella* spp.**CATALOG NUMBER**

V-PAT04

ORIGINAL CERTIFICATION DATE

July 2, 2026

PRINCIPLE OF THE METHOD

The Elevia 1.1 *Salmonella* spp. kit is a real-time RT-PCR kit designed for the detection of RNA from bacteria of the genus *Salmonella* in surface samples, enriched cocoa-based products, and other low water activity foods. Detection of *Salmonella* spp. is carried out by amplification of specific nucleic acids extracted using the Magneus 1 Bacteria (food samples) or Magneus 6 Bacteria (surface samples) extraction kits. The genetic material is reverse-transcribed, and then amplified by real time PCR, using specific fluorescent probes for detection. The Elevia 1.1 *Salmonella* spp. kit has a detection limit of 5 copies/ μ L and a sensitivity of 1 cfu/375 g for food samples after enrichment and 1 cfu/100 cm² for surface samples after enrichment. The Elevia 1.1 *Salmonella* spp. kit is compatible with real-time PCR instruments equipped with FAM and HEX detection channels.

CERTIFIED CLAIM STATEMENT: The Elevia 1.1 *Salmonella* spp. method is certified for the detection of *Salmonella* spp. within the scope of Tables 1 and 2.

Certified method includes:

1. Bio-Rad CFX96 Deep Well Real-Time PCR System
2. Bio-Rad CFX Opus Standard Real-Time PCR System

Table 1. Method performance claims

Matrix	Test portion	Enrichment conditions				Reference method ^b	Claim ^c
		Broth ^a	Volume	Temperature	Time		
62% cocoa chocolate	375 g	Enhanced Augmentis 91 BPW	3375 mL	40 $\pm$ 1°C	10-24 h	ISO 6579-1:2017	NSDD
Cocoa liquor	375 g	Enhanced Augmentis 91 BPW	3375 mL	40 $\pm$ 1°C	10-24 h	ISO 6579-1:2017	NSDD
Non-fat dry milk	375 g	BPW-ISO	3375 mL	37 $\pm$ 1°C	8-24 h	ISO 6579-1:2017	NSDD
Peanuts	375 g	BPW-ISO	3375 mL	37 $\pm$ 1°C	8-24 h	ISO 6579-1:2017	NSDD
Stainless steel	100 cm ² , swab ^d	Augmentis Xpress 1	2.2 mL	37 $\pm$ 1°C	3-24 h	ISO 6579-1:2017	NSDD
	100 cm ² , sponge ^e	Augmentis Xpress 1	30 mL	37 $\pm$ 1°C	3-24 h	ISO 6579-1:2017	NSDD

^a Enhanced Augmentis 91 BPW = Augmentis 91 BPW + Potentia 1 *Salmonella* (TAAG); BPW-ISO = Buffered Peptone Water, ISO formulation

^b ISO = International Organization for Standardization; includes ISO 6579-1:2017/Amd.1:2020

^c NSDD = No statistical difference detected using SLV study design from OMA Appendix J (2012). The SLV qualitative method comparison study design from OMA Appendix J (2012) is not intended to demonstrate statistical equivalence. Expert opinion is that the method is appropriate for its intended use.

^d Swabs were validated premoistened in Augmentis Xpress 1 broth and Lethen neutralizing broth.

^e Sponges were validated premoistened in Lethen neutralizing both.

Table 2. Method selectivity

Broth ^a	Temperature	Inclusivity species		Exclusivity species	
		No. tested	No. positive	No. tested	No. positive
Enhanced Augmentis 91 BPW	35–38°C	102 ^b	102	40 ^c	0

^a Enhanced Augmentis 91 BPW = Augmentis 91 BPW + Potentia 1 *Salmonella* (TAAG)

^b Comprising *Salmonella bongori* and *Salmonella enterica* including subsps. *arizonae*, *diarizonae*, *indica*, *houtenae*, *salamae*, and *enteric*

^c Comprising 39 species

Table 3. Method history

No.	Date	Summary	Supporting data
1	July 2, 2026	Original Certification	Certification Report